Cryptosporidium varanii infection in leopard geckos (Eublepharis macularius) in Argentina

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Abstract
Cryptosporidiosis is observed in reptiles with high morbidity and considerable mortality. The objective of this study was to achieve the molecular identification of Cryptosporidium spp. in pet leopard geckos (Eublepharis macularius) from a breeder colony in Buenos Aires, Argentina. Oocysts comparable to those of Cryptosporidium spp. were detected in three geckos with a history of diarrhea, anorexia and cachexia. Molecular identification methods confirmed the presence of Cryptosporidium varanii (syn. C. saurophilum). This agent was considered to be the primary cause of the observed clinical disease. This is the first description of C. varanii infection in pet reptiles in Argentina.

Keywords: Argentina, Cryptosporidium spp., Leopard gecko (Eublepharis macularius), PCR, Sequencing.

Introduction
Cryptosporidiosis infection (referred as cryptosporidiosis) is a zoonosis of worldwide distribution that affects the gastrointestinal tract of a range of vertebrate hosts, including mammals, reptiles, birds and fish (Fayer, 2010). Cryptosporidiosis in humans is a cause of diarrhea, mainly in children, and could cause a severe disease in immunocompromised patients (Xiao et al., 2004a). Cryptosporidiosis has been described in many different reptile species, and is especially important in snakes and lizards. Two main Cryptosporidium species have been identified in reptiles: C. serpentis and C. varanii (syn. C. saurophilum). C. serpentis is a gastric parasite found mainly in snakes, and frequently associated with prominent clinical signs like anorexia, postprandial regurgitation, lethargy, mid-body swelling, and weight loss, while infections in lizards are usually asymptomatic (Fayer et al., 1997). C. varanii is an intestinal parasite found primarily in lizards (Pavlasek and Ryan, 2008; Xiao et al., 2004b) and can cause anorexia, progressive weight loss, abdominal swelling and high mortality, particularly in juvenile lizards (Koudela and Modry, 1998). Other Cryptosporidium spp. have been described in reptiles like C. muris, C. parvum, C. parvum mouse genotype (syn. C. tyzzeri). However, these Cryptosporidium spp. oocyst could represent a passage of parasites from ingested prey or feeder mice (Xiao et al., 2004b).

Reptiles are becoming popular pets worldwide; however, little is known about the presence and control of cryptosporidiosis in this animal population. In the last several years, cryptosporidiosis has caused important economic losses for commercial reptile breeders. Therefore, the objective of this study was to perform molecular characterization of Cryptosporidium spp. from naturally infected leopard geckos (Eublepharis macularius) in Argentina.

Materials and Methods

Animals and samples
A pooled stool sample from 3 leopard geckos (E. macularius) was collected and submitted to La Plata University for molecular studies. Three geckos (two females and 1 male) (Fig. 1) were received by veterinary practitioners from a breeder colony in Buenos Aires, Argentina with a history of diarrhea, anorexia and cachexia. Faecal samples were first placed in a 15-ml tube. The tube was partially filled with sucrose solution and mixed in a vortex for approximately 5 sec each tube was centrifuged at 252 g for 10 min. An aliquot was taken from the surface and spread on a slide and examined for the presence of Cryptosporidium spp. oocysts by light microscopy (Deming et al., 2008). The modified Ziehl-Neelsen stain for fecal smears was performed essentially as described previously (Henriksen and Pohlenz, 1981); were evidenced several round oocysts of 4-5 μm in diameter compatible with Cryptosporidium spp. (Fig. 2). The pooled sample was subjected to a sugar flotation technique (sucrose solution...
without formaldehyde) and water sedimentation as described previously for concentration of coccidian oocysts (Ortega-Mora et al., 2007). Concentrated oocysts were re-suspended in 450 μl of nuclease free water.

**DNA extraction**

Oocyst disruption and DNA purification from the faecal sample (processed in triplicate) was performed with a DNA stool commercial kit (QIAtip DNA Stool Mini Kit, Qiagen, Hilden, Germany) according to the manufacturer’s recommendations. A process control sample (extraction kit solutions and 150 μl of nuclease free water) was evaluated together with the stool samples.

**Molecular identification**

A nested PCR technique was performed for amplification of a polymorphic fragment of the 18S rDNA gene (Xiao et al., 1999). Five μl aliquot of the internal PCR products was examined by electrophoresis in 1% agarose gels and stained with ethidium bromide. Two of the amplicons obtained (which reach a gel estimated concentration of about 40 ng/μl), were purified using polyethylene glycol and sequenced in both directions using the Megabace 1000 Sequencing (GE Health care) at IGEVET, FCV, UNLP. Sequences were aligned and analyzed using GENEIOUS software (free available version 7.1, http://www.geneious.com). Consensus sequences obtained were compared with others published in GenBank by BLASTn megablast analysis.

**Results and Discussion**

Reptiles have become popular pets worldwide in recent years. The leopard gecko is a crepuscular ground-dwelling lizard naturally found in the deserts of Asia and throughout Pakistan, to the northwestern parts of India (Henkel et al., 1995).

Upon ingestion of Cryptosporidium spp. oocysts by the host, sporozoites are released and invade epithelial cells. The protozoans multiply asexually and sexually causing death of the host cells. Cryptosporidium spp. infection is associated with hyperplastic and inflammatory lesions of the gastrointestinal tract in different animal species including geckos (Terrell et al., 2003). Thus, Cryptosporidiosis has been associated with a wasting syndrome or “going light” in leopard geckos, characterized by chronic weight loss, diarrhea, lethargy, followed by anorexia and death (Deming et al., 2008). In the present study, three leopard geckos from a breeder colony in Argentina showed a wasting syndrome and oocysts compatible with Cryptosporidium spp. were detected by Ziehl-Neelsen staining. Previous studies highlight the importance of molecular diagnostic methods to identify species level within the genus Cryptosporidium (Richter et al., 2011). In Spain, Pedraza-Diaz et al. (2009) performed PCR and sequencing from seven leopard geckos stool samples with Cryptosporidium spp. oocysts, identifying C. varanii (n = 3), C. serpentis (n = 2) and C. parvum (n = 2). A similar study performed in Austria, detected C. varanii and C. serpentis (32/74 and 8/74 leopard geckos, respectively) (Richter et al., 2011).

In the present study, molecular identification of Cryptosporidium spp. was performed by sequencing of a polymorphic fragment of the 18s rRNA gene. The nested PCR was positive (all 3 replicates), evidencing a product of around 830 bp. A consensus sequence of 748 bp was obtained and registered on GenBank under accession number KM610237. This sequence was compared with others published in GenBank by BLASTn megablast analysis and revealed a 99% sequence identity with C. saurophilum (GenBank EU555551, EU555552 and EF502042) from asymptomatic leopard geckos (E. macularius) and one snake (Boa constrictor) in Spain (Pedraza-Diaz et al., 2009), and from symptomatic a adult snake...
(Elaphe guttata guttata) from a private snake breeding colony from Japan (Plutzer and Karanis, 2007), respectively. The sequence evidenced similar identity (99%) with *C. varanii* (GenBank KJ000485) from an asymptomatic adult snake (*Pantherophis guttatus*) from the serpentarium of the Butantan Institute in São Paulo, Brazil (da Silva et al., 2014), and *Cryptosporidium* spp. from a desert monitor (GenBank AF112573) from Missouri, USA (Xiao et al., 1999).

*C. varanii* was initially named in 1995 by Pavlásek et al. (1995) to describe oocysts obtained from an Emerald monitor (*Varanus prasinus*) in the Prague Zoo. The description was based on oocyst morphology, histology of endogenous stages in the intestine, and failure to infect mice. The same species was subsequently identified in other lizards and in snakes (Pavlasek and Ryan, 2008). The nomenclature *C. saurophilum* was used in 1998 by Koudela and Modry (1998) for oocysts obtained from a Schneider’s skink (*Eumeces schneideri*). Molecular comparison between *C. saurophilum* and *C. varanii* at the 18S RNA and actin loci showed that they are genetically identical, but as *C. varanii* was described previously, it takes precedence over *C. saurophilum* which should be considered a junior synonym of *C. varanii* (Pavlasek and Ryan, 2008; Xiao et al., 2004b). Considering the previous studies, the detected protozoan from leopard geckos in Argentina was identified as *C. varanii* (syn. *C. saurophilum*).

*Cryptosporidium* spp. obtained from a desert monitor was originally reported by Xiao et al. (1999). Morphologically, oocysts of the *Cryptosporidium* spp. desert monitor genotype were very similar to those of *C. varanii* in shape and size and significantly smaller than oocysts of *C. serpentis*. Molecular and biologic characterizations conducted in later studies identified the parasite as *C. saurophilum* (Xiao et al., 2004b). This could explain why the sequence obtained in the present study evidenced 99% homology with the sequence GenBank AF112573 (Xiao et al., 1999). The sequence names reported in GenBank should be revised in order to avoid further misinterpretations.

This is the first case report and molecular identification of *C. varanii* in leopard geckos in Argentina. Even though it was not possible to determine whether all three animals were infected with *C. varanii* since a pooled sample was received in the laboratory, it is possible to assume that all animals were infected by *C. varanii* as they all evidenced similar clinical signs compatible with cryptosporidiosis. Further studies should be conducted to identify the prevalence and implications of cryptosporidiosis in pet lizards from Argentina in order to generate awareness among commercial breeders, pet owners and clinical practitioners in Argentina. The present study evidenced the existence of clinical cryptosporidiosis in captive leopard geckos in Argentina and highlights the importance of molecular identification to species level of *Cryptosporidium* spp.-like oocysts found in fecal samples.

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**Conflict of interest**

The authors declare that there is no conflict of interest.

**References**


